Mark J. Snider Robert E. Wilson Professor of Chemistry

Department of Chemistry The College of Wooster 931 College Mall Wooster, OH 44691

https://www.wooster.edu/bios/msnider/

(330) 263-2391 msnider@wooster.edu Google Scholar Profile ORCiD: 0000-0003-1054-1276 Scopus Author ID: 7007076689

EDUCATION	Ph.D. 2001	Biochemistry & Biophysics University of North Carolina – Chapel Hill Advisor: Professor Richard Wolfenden Dissertation title: Thermodynamic Analysis of the Mechanism of Escherichia coli Cytidine Deaminase © 2001
	B.A. 1997	Chemistry Summa Cum Laude Capital University, Columbus Ohio
PROFESSIONAL APPOINTMENTS	2015 – 2013 – 2012 2007 – 2008 2007 – 2015 2002 – 2007 2001 – 2002	Professor of Chemistry, The College of Wooster Robert E. Wilson Endowed Professorship in Chemistry Honorary Fellow, Department of Biochemistry, University of Wisconsin-Madison (sabbatical) Visiting Scientist, Department of Chemistry & Chemical Biology, Cornell University (sabbatical) Associate Professor of Chemistry, The College of Wooster Assistant Professor of Chemistry, The College of Wooster Postdoctoral Teaching/Research Fellow, Department of Chemistry, The College of Wooster (Mentor: Prof. Charles L. Borders, Jr.)
RESEARCH LEAVES	2018 – 2019; 2023 2012 – 2013 2007 – 2008	at The College of Wooster; Transient state kinetic analysis of the mechanism of 6-hydroxynicotinate-3-monooxygenase, and the genes of nicotinate degradation in <i>Bacillus niacini</i> . with Professor W. W. Cleland, Department of Biochemistry, University of Wisconsin, Madison, WI and with Professor David Ballou, Department of Biological Chemistry, University of Michigan, Ann Abor, MI with Professor Tadhg P. Begley, Department of Chemistry & Chemical Biology, Cornell University, Ithaca, NY
CURRENT RESEARCH	degradation; investigate the of phosphage	nd structural enzymology of bacterial nicotinic acid (vitamin B ₃) Transient-state kinetics by stopped-flow spectrophotometry to e mechanisms of flavin monooxygenases; Functional characterization n kinases to understand molecular evolution of cooperative ligand uaternary structure.

EXTERNALLY-FUNDED RESEARCH GRANTS

2018-2023 National Science Foundation: Molecular Biophysics Program

RUI: Collaborative Research: Enzymology of Bacterial Nicotinic Acid Catabolism by Mark Snider (PI, Wooster), Katherine Hicks (PI, SUNY Cortland) \$ 551,536

2016-2019: National Science Foundation: Major Research Instrumentation Program

Acquisition of an NMR Spectrometer to Sustain Excellence in Undergraduate Research by Paul Bonvallet (PI), Judith Amburgey-Peters (Co-PI), Spring Knapp (Co-PI), Mark

Snider (Co-PI) and Sarah Sobeck (Co-PI); \$343,697

2012-2014: Research Corporation for Science Advancement: Multi-Investigator Cottrell College

Science Award Program: Bacterial Degradation of Pharmaceuticals and Personal Care Products During Waste Water Treatment; by Stephanie Strand (Microbiologist) and Melissa Schultz (Analytical Chemist) and Mark Snider (Biochemist); \$100,000

2008-2011: National Science Foundation: Major Research Instrumentation Program

Acquisition of an LC/MS/MS to Enhance Undergraduate Research and Teaching by Melissa Schultz (PI), Paul Edmiston (Co-PI) and Mark Snider (Co-PI); \$ 226,115

2006-2009: National Science Foundation: Major Research Instrumentation Program

Acquisition of an Isothermal Titration Calorimeter

by Paul Bonvallet (PI), Don Jacobs (Co-PI) and Mark Snider (Co-PI); \$ 91,090

2004-2007: National Science Foundation: Metabolic Biochemistry Program

RUI: Comparative Structure – Function Analysis of Phosphagen Kinases by Mark Snider (PI), Paul Edmiston (Co-PI) and Dean Fraga (Co-PI); \$ 320,307

PEER-REVIEWED PUBLICATIONS (*College of Wooster undergraduate) *Manuscripts in progress:*

M. J. Snider, I. Lee, M. Aryal*, I. Eccles-James*, J. Anquandah*, J. Fishovitz, J. Graham*, R. Klein*, L. Marcze*, L. Stetzik*, G. Kerwood and D. Fraga; Evolution of inter-subunit communication in the phosphagen kinases: an alternating sites reactivity mechanism for catalysis; *manuscript in progress– to be submitted to Biochemistry.*

P. Chiaranunt*, E. D. Sullivan*, N. Spittle*, M. Anderson, K. Morrison, D. Hilmey, R. S. Rowlett, and M. J. Snider; A structural and functional investigation of the catalytic role of Cys150-Asp29-Lys117 in maleamate amidohydrolase – manuscript in progress, to be submitted to Biochemistry.

J. Redick*, M. Hlaing*, R. Campbell*, L. Rajakovich, and M. J. Snider (2024) Mechanistic insights into substrate ionization and decarboxylation by 6-hydroxynicotinate 3-monooxygenase with 5-Cl-6-hydroxynicotinate, an efficient substrate analogue – *manuscript in progress, to be submitted to Biochemistry.*

Manuscripts published: (*College of Wooster undergraduate)

- 32. B. C. Richardson, Z. R. Turlington, S. V. Ferreira de Macedo, S. K. Phillips, K. Perry, S. G. Brancato, E. W. Cooke*, J. R Gwilt, M. Dasovich*, A. J. Roering, F. M. Rossi, M. J. Snider, J. B. French and K. A. Hicks (2024) Structural and functional characterization of a novel class A flavin monooxygenase from *Bacillus niacini*. *Biochemistry 63: 2391-2532*. <u>DOI</u>
- 31. Z. R. Turlington; S. V. Ferreira de Macedo; K. Perry; S. L. Belsky*; J. A. Faust; M. J. Snider and K. A. Hicks (2024) Ligand bound structure of a 6-hydroxynicotinic acid 3-monooxygenase provides mechanistic insights; *Archives in Biochemistry & Biophysics 752*; 109859. *DOI*

- S. K. Phillips, S. G. Brancato, S. N. MacMillan, M. J. Snider, A. J. Roering and K. A. Hicks (2023) Synthesis, crystallographic and spectroscopic characterization of 2,6-dihydroxypyridine; Acta Crystallographica Section E79. DOI
- 29. S. W. Perkins*, M. Z. Hlaing*, K. A. Hicks, L. Rajakovich, and M. J. Snider (2023) Mechanism of the multistep catalytic cycle of 6-hydroxynicotinate 3-monooxygenase revealed by global kinetic analysis; *Biochemistry 62;* 1553-1567. DOI
- 28. P. Piper*, B. Begres*, M. Snider, and D. Fraga (2020) Two *cryptosporidia* species encode active creatine kinases that are not seen in other *Apicomplexa* species; *Comparative Biochemistry and Physiology Part B 246*, 110459. DOI
- 27. D. Fraga., K. Stock*, M. Aryal*, C. Demoll*, L. Fannin*, and M. J. Snider (2019) Bacterial arginine kinases have a highly skewed distribution within the Proteobacteria. *Comparative Biochemistry and Physiology Part B* 233, 60-71. DOI
- 26. K. D. Nakamoto*, S. W. Perkins*, R. G. Campbell*, M. R. Bauerle*, T. G. Gerwig*, S. Gerislioglu, C. Wesdemiotis, M. A. Anderson, K. A. Hicks and M. J. Snider (2019) Mechanism of 6-hydroxynicotinate 3-monoxygenase, a flavin-dependent decarboxylative hydroxylase involved in aerobic nicotinic acid degradation. *Biochemistry 58*; 1751-1763. DOI
- 25. K. A. Hicks, M. E. Yuen, W.- F. Zhen, T. J. Gerwig*, R. W. Story*, M. Kopp, and M. J. Snider (2016) Structural and biochemical characterization of 6-hydroxynicotinic acid 3-monooxygenase, a novel decarboxylative hydroxylase involved in aerobic nicotinate degradation. *Biochemistry 55*, 3432–3446.
- 24. D. Fraga, M. Aryal*, J. E. Hall*, E. Rae*, and M. Snider (2015) Characterization of the arginine kinase isoforms in *Caenorhabditis elegans*. *Comparative Physiology and Biochemistry; Part B: Biochemistry & Molecular Biology 187*; 85-101. DOI
- 23. Z. H. Harvey* and M. J. Snider (2014) Draft genome of the nicotinate-metabolizing soil bacterium *Bacillus niacini* (DSM 2923) *Genome Announc. 2* (6) e01251-14. DOI
- 22. A. Palmer*, B. Begress*, J. Van Houten*, <u>M. J. Snider</u>, and D. Fraga **(2013)** Characterization of a putative oomycete taurocyamine kinase: Implications for the evolution of the phosphagen kinase family.

 Comparative Physiology and Biochemistry; Part B: Biochemistry & Molecular Biology, 166; 173-181. <u>DOI</u>
- 21. G. K. Schroeder, L. Zhou, M. J. Snider, X. Chen, and R. Wolfenden (**2012**) The flight of a cytidine deaminase complex with an imperfect transition state analogue inhibitor: mass spectrometric evidence for the presence of a trapped water molecule. *Biochemistry 51*, 6476-6486. **DOI**
- 20. J. Bragg, A. Rajkovic*, C. Anderson, R. Curtis, J. V.-Houten*, B. Begres*, C. Naples*, <u>M. Snider</u>, D. Fraga and M. Singer (**2012**) Identification and characterization of an arginine kinase from *Myxococcus xanthus* required for fruiting body formation and cell differentiation; *J. Bacteriology 194*, 2668-2676. <u>DOI</u>
- 19. V. A. Kincaid*, E. D. Sullivan*, R. D. Klein*, Jeff W. Noel*, R. S. Rowlett, M. J. Snider (**2012**) Structure and catalytic mechanism of nicotinate (vitamin B₃) degradative enzyme maleamate amidohydrolase from *Bordetella Bronchiseptica RB50*; *Biochemistry 51*, 545-554. DOI
- 18. A. Hazra, M. J. Snider, A. Chatterjee, D. Chatterjee, J. W. Hanes, D. G. Hilmey, K. Krishnamoorthy,

- K. M. Mccullock, S. O'Leary, J. M. Sanders, M. J. Waitner, and T. P. Begley. Coenzyme and prosthetic group biosynthesis; *Encyclopedia of Microbiology;* Moselio Schaechter, Ed. (**2009**) Vol. 5, 79-88, Oxford: Elsevier. <u>Link</u>
- 17. L. D. Andrews*, J. Graham*, <u>M. J. Snider</u>, D. Fraga. (**2008**) Characterization of a novel bacterial arginine kinase from *Desulfotelea psychrophilia Comparative Physiology and Biochemistry; Part B: Biochemistry & Molecular Biology 150:* 312-319. <u>DOI</u>
- 16. M. J. Jourden*, C. N. Clarke*, A. K. Palmer*, E. J. Barth, R. C. Prada*, R. N. Hale*, D. Fraga, M. J. Snider, P. L. Edmiston (2007) Changing the substrate specificity of creatine kinase from creatine to glycocyamine: evidence for a highly evolved active site. *Biochimica et Biophysica Acta Proteins and Proteomics 1774*: 1519-1527. DOI
- 15. M. J. Jourden*, M. J. Thomenius*, P. R. Geiss*, L. A. Horst*, M. M. Barty*, G. B. Mulligan*, R. M. Almeida*, B. A. Kersteen*, N. R. Myers*, M. J. Snider, C. L. Borders, Jr., P. L. Edmiston (2005)

 Transition state stabilization by six arginines clustered in the active site of creatine kinase. *Biochimica et Biophysica Acta Proteins and Proteomics 1751*: 178-183. DOI
- 14. C. H. Borchers, V. E. Marquez, G. K. Gottfried, S. E. Short, M. J. Snider, P. Speir and R. Wolfenden (**2004**) Fourier transform ion cyclotron resonance mass spectrometry reveals the presence of a water molecule in an enzyme-transition state analogue complex. *Proceedings of the National Academy of Sciences USA 101:* 15341-15345. **DOI**

Review

- 13. <u>M. Snider</u>, B. Temple and R. Wolfenden (**2004**) The path to the transition state in enzyme reactions: a survey of catalytic efficiencies. *Journal of Physical Organic Chemistry 17:* 586-591. <u>DOI</u>
- 12. C. L. Borders, Jr., K.M. MacGregor*, P. L. Edmiston, E. R. K. Gbeddy*, M. J. Thomenius*, G. B. Mulligan*, and M. J. Snider (2003) Asparagine 285 plays a key role in transition state stabilization in rabbit muscle creatine kinase. *Protein Science* 12: 532-537. DOI
- 11. J.M. Cox*, C.A. Davis*, C. Chan*, M.J. Jourden*, A.M. Jorjorian*, M.J. Brym*, <u>M.J. Snider</u>, C.L. Borders, Jr., P.L. Edmiston (**2003**) Generation of an active site monomer of rabbit muscle creatine kinase by site-directed mutagenesis: the effect of quaternary structure on catalysis and stability. *Biochemistry 42*: 1863-1871. <u>DOI</u>
- 10. C. L. Borders, Jr., M. J. Snider, R. Wolfenden, and P. L. Edmiston (**2002**) Determination of the affinity of each component of a composite quaternary transition-state analog complex of creatine kinase. *Biochemistry 41*: 6995-7000. DOI
- 9. <u>M. J. Snider</u>, D. Lazarevic, and R. Wolfenden (**2002**) Catalysis by entropic effects: the action of cytidine deaminase on 5,6-dihydrocytidine. *Biochemistry 41*: 3925-3930. <u>DOI</u>
- 8. <u>M. J. Snider</u>, L., Reinhardt, R. Wolfenden, and W. W. Cleland (**2002**) ¹⁵N kinetic isotope effects on uncatalyzed and enzymatic deamination of cytidine. *Biochemistry* 41: 415-421. **DOI**

Review

- 7. R. Wolfenden and M. J. Snider (2001) The depth of chemical time and the power of enzymes as catalysts. *Accounts of Chemical Research 34:* 938-945. DOI
- 6. <u>M. J. Snider</u> and R. Wolfenden (**2001**) Site-bound water and the shortcomings of a less-than-perfect transition state analogue. *Biochemistry 40:* 11364 11371. <u>DOI</u>

- 5. B. G. Miller, M. J. Snider, R. Wolfenden, and S. A. Short. (2001) Dissecting a charged network at the active site of orotidine 5'-monophosphate decarboxylase. *Journal of Biological Chemistry 276*:15174-15176. DOI
- 4. <u>M. J. Snider</u> and R. Wolfenden (**2000**) The rate of spontaneous decarboxylation of amino acids. *Journal of the American Chemical Society 122*: 11507-11508. <u>DOI</u>
- 3. M. J. Snider, S. Gaunitz, C. Ridgway, S. A. Short, and R. Wolfenden (**2000**) Temperature effects on the catalytic efficiency, rate enhancement, and transition state affinity of cytidine deaminase, and the thermodynamic consequence for catalysis of removing a substrate 'anchor'. *Biochemistry 39*: 9746-9753. **DOI**
- 2. B. G. Miller, M. J. Snider, S. A. Short, and R. Wolfenden (**2000**) Contribution of enzyme-phosphoryl contacts to catalysis by orotidine 5'-phosphate decarboxylase. *Biochemistry 39*: 8113-8118. **DOI**
- 1. R. Wolfenden, <u>M. Snider</u>, C. Ridgway, and B. Miller (**1999**) The temperature dependence of enzyme rate enhancements. *Journal of the American Chemical Society 121*: 7419-7420. <u>DOI</u>

PRESENTATIONS AT PROFESSIONAL MEETINGS (*College of Wooster Undergraduate; presenter)

<u>Aglesia Aregawi</u> and Mark J. Snider (**2025**) Investigating Substrate Promiscuity of 6-Hydroxynicotinate 3-monooxygenase (NicC): NicC Activity with 5-Cl-Coumalate; *American Society of Biochemistry & Molecular Biology National Meeting*, Chicago, IL.

<u>Julian Pac</u> and Mark J. Snider (**2025**) Unraveling the Cooperative Dynamics of PhzS and PhzM: Understanding the Ordered Steps in Pyocyanin Biosynthesis by *Pseudomonas aeruginosa; American Society of Biochemistry & Molecular Biology National Meeting*, Chicago, IL.

<u>Mayank Pandey</u> and Mark J. Snider (2025) Role of Arg108 in the Binding of NADH by 6-Hydroxynicotinate 3-monooxygenase (NicC); *American Society of Biochemistry & Molecular Biology National Meeting*, Chicago, IL. [Honorable Mention for Best Poster in Enzymology Section of the ASBMB National Undergraduate Poster Competition]

<u>Charitha Patlolla</u> and Mark J. Snider (**2025**) Role of Arg184 in substrate binding and coenzyme coordination by 6-hydroxynicotinate 3-monooxygenase (NicC); *American Society of Biochemistry & Molecular Biology National Meeting*, Chicago, IL. [Honorable Mention for Best Poster in Enzymology Section of the ASBMB National Undergraduate Poster Competition]

<u>Sipara Semu</u> and Mark J. Snider (**2025**) Role of Conserved Arginines in Substrate Binding and Flavin Dynamics in 6-Hydroxynicotinate Monooxygenase; *American Society of Biochemistry & Molecular Biology National Meeting*, Chicago, IL.

<u>Mark J. Snider</u> and Katherine A. Hicks (**2024**) Novel enzymology of nicotinic acid degradation in *Bacillus niacini*. <u>28th Enzyme Mechanisms Conference</u>, Naples, FL, January 2024.

<u>Mark J. Snider</u>, Jack Redick, May Hlaing and Lauren Rajakovich (**2023**) Probing substrate specificity in the class A flavin enzyme 6-hydroxynicotinate 3-monooxygenase; <u>Midwest Enzyme Chemistry Conference</u>, Northwestern University, Evanston, IL, September 2023.

<u>Takoda T. Zuehlke*, Jack Donahue</u>*, and Mark J. Snider (**2023**) Identification of a Putative Binding Mechanism for the Reducing Substrate of 6-Hydroxynicotinate 3-Monooxygenase; *American Society of*

Biochemistry & Molecular Biology National Meeting, Seattle, WA. [Honorable Mention for Best Poster in Enzymology Section of the ASBMB National Undergraduate Poster Competition]

<u>Eric Senanu Adadevoh*</u> and Mark J. Snider (**2023**) Exploring the Binding Multiplicity, Mode of Inhibition and Communication between NADH Binding Sites on 6-Hydroxynicotinate 3-Monooxygenase (NicC); American Society of Biochemistry & Molecular Biology National Meeting, Seattle, WA.

<u>Kira Boyce*, Katherine Olson*</u>, and Mark J. Snider (**2023**) A Critical Role for a *CoxG* Homologue in the Function of *B. Niacini* Nicotinate Dehydrogenase; *American Society of Biochemistry & Molecular Biology National Meeting*, Seattle, WA.

<u>Sam L. Belsky</u>*, <u>Sipara H. Semu</u>*, Mark J. Snider, Zachary R. Turlington, and Katherine A. Hicks (**2023**) Defining the Role of A Conserved Motif at the Interface of the Substrate and FAD binding sites in FAD dynamics in class A flavin Monooxygenases: A Case Study of 6-Hydroxynicotinate-3-Monooxygenase; *American Society of Biochemistry & Molecular Biology National Meeting*, Seattle, WA.

Robby Beal*, Merlin Li*, Dean Fraga and Mark J. Snider (2022); Determining whether tightness of binding in the active site of a dimeric taurocyamine kinase plays a role in inducing negative cooperativity; American Society of Biochemistry & Molecular Biology National Meeting, FASEB J 36, S1. [Honorable Mention for Best Poster in Protein Structure/Function and Enzymology Section of the ASBMB National Undergraduate Poster Competition]

<u>Sam Belsky</u>* and Mark J. Snider (**2022**) Exploring the promiscuity potential of 6-hydroxynicotinate-3-monooxygenase: consequences to catalysis of adding a nitrogen at C5 within the substrate's aromatic ring; *American Society of Biochemistry & Molecular Biology National Meeting*, FASEB J 36, S1, R2629.

<u>Joel Brown*</u> and Mark J. Snider (**2022**) Effector or substrate: characterizing the molecular features of 6-HNA that govern enhanced rates of NADH oxidation and hydroxylation by 6-hydroxnicotinate-3-monoxygenase (NicC); *American Society of Biochemistry & Molecular Biology National Meeting*, <u>FASEB J 36</u>, S1, R3990. [Winner, Best Poster Presentation in Protein Structure/Function and Enzymology Section of the ASBMB National Undergraduate Poster Competition].

<u>Emilee Haines*, Zoë Semersky*</u>, Luke Borgelt*, and Mark J. Snider (**2022**); Determination and Characterization of Genes that Encode the Nicotinate Dehydrogenase and 6-Hydroxynicotinate Dehydrogenase Complexes within the Nicotinic Acid Degradation Pathway by *Bacillus niacini; American Society of Biochemistry & Molecular Biology National Meeting, FASEB J* 36, S1.

<u>May Z. Hlaing</u>*, Ryan Campbell*, and Mark J. Snider (**2022**); Investigating substrate specificity for 6-hydroxynicotinate 3-monooxygenase (NicC) With coumalic acid: consequences to catalysis of replacing or removing the ring nitrogen. *American Society of Biochemistry & Molecular Biology National Meeting*, <u>FASEB J 36, S1, R3597.</u>

<u>Sai K. Khal</u>*, Noah Helton* and Mark J. Snider (**2022**) Elucidating the nicotinic acid degradation pathway in *Bacillus niacini*: Biochemical Characterization of proteins of unknown function; *American Society of Biochemistry & Molecular Biology National Meeting*, *FASEB J* 36, S1.

Merlin Li*, Dean Fraga, and Mark J. Snider; Evidence of negative cooperativity in cytosolic taurocyamine kinase from *Arenicol brasiliensis* and its implicatdions in the evolution of the phosphagen kinase family. 10th Annual Southeast Enzyme Conference, Virtual, April **2021**. <u>Mark J. Snider</u>, Scott W. Perkins*, Ryan G. Campbell*, and Lauren Rajakovich; Critical role for substrate ionization in the mechanism of 6-hydroxynicotate 3-monooxygenase. *Gordon Research Conference on Enzymes, Coenzymes and Metabolic Pathways*, Waterville Valley, NH, July **2019**.

<u>Scott W. Perkins* and Mark J. Snider</u>; Mechanism of 6-hydroxynicotinate 3-monooxygenase (NicC), a flavin-dependent decarboxylative hydroxylase involved in aerobic nicotinic acid catabolism. *26th Enzyme Mechanisms Conference*. New Orleans, LA, January **2019**.

<u>Katherine A. Hicks,</u> Tariq Hussain, Scott W. Perkins*, Kent Nakamoto*, and Mark J. Snider; Characterization of the catalytic mechanism of NicC, a 6-hydroxynicotinic acid 3-monooxygenase using active site variants. *Gordon Research Conference on Enzymes, Coenzymes and Metabolic Pathways*, Waterville Valley, NH, July **2018**. [Selected for an oral presentation]

<u>Scott W. Perkins*</u> and Mark J. Snider (**2018**) Investigating putative key catalytic residues and uncoupled hydroperoxyflavin formation in the mechanism of 6-hydroxynicotinate-3-monooxygenase, a decarboxylative-hydroxylase in bacterial nicotinate catabolism; *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **32**:655.13 [Honorable Mention for Best Poster in Metabolism and Bioenergetics Section of the ASBMB National Undergraduate Poster Competition]

<u>Kent Nakamoto*</u>, Selim Gerislioglu, and Mark J. Snider (**2018**) Determining the mechanism of 6-hydroxynicotinate 3-monooxygenase (NicC), an enzyme involved in nicotinate degradation; *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **32**:655.14

<u>Nathan Brownstein*</u>, and Mark J. Snider (**2018**) Expression and functional characterization of active nicotinic acid dehydrogenase from *Pseudomonas fluorescens* Pf5; *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **32**:796.8

<u>Sydney Fine*</u>, and Mark J. Snider (**2018**) Elucidating the Role of a Putative Monooxygenase in the Catabolism of Nicotinic Acid and Nicotine by *Bacillus niacini; American Society of Biochemistry & Molecular Biology National Meeting; FASEB J 32:536.4*

<u>Kaeli Zoretich,*</u> Sydney Fine*, and Mark J. Snider (**2018**) The Catabolism of Nicotinic Acid and Nicotine and the Role of a Putative Monooxygenase in *Bacillus niacini; American Society of Biochemistry & Molecular Biology National Meeting; FASEB J 32:536.6*

<u>Kent Nakamoto*, Scott W. Perkins*,</u> and Mark J. Snider (**2018**) Determining the mechanism of 6-hydroxynicotinate 3-monooxygenase (NicC). Meeting-in-Miniature, Cleveland Section of the American Chemical Society, Oberlin College, March 2018.

<u>Scott W. Perkins*</u>, Kent Nakamoto* and Mark J. Snider (**2017**) Investigating the role of covalent intermediates in the mechanism of 6-hydroxynicotinic acid 3-monoxygenase, a decarboxylative-hydroxylase in bacterial nicotinate catabolism; *Midwest Enzyme Chemistry Conference*, Loyolla University of Chicago, October 2017.

<u>Kathleen Kalafatis*</u> and Mark J. Snider (**2017**) Characterization of the nicotinic acid and 6-hydroxynicotinic acid dehydrogenase complexes in *Bacillus niacini; American Society of Biochemistry & Molecular Biology National Meeting; FASEB J 31:919.3*

<u>Tsun Ki Jerrick To*</u> and Mark J. Snider (**2017**) Genetic and functional characterization of the enzyme of nicotinic acid degradation in *Bacillus niacini*. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 31:624.6**

<u>Laura Sherer*</u>, Mark Snider, Maria_Eirini Pandelia, and Roger Rowlett (**2017**) Cofactor analysis of nicotinate dehydrogenase and 6-hydroxynicotinate dehydrogenase in *Bacillus niacini*. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 31:919.4**

<u>Mark J. Snider</u> and Katherine Hicks; Structural and mechanistic studies of 6-hydroxynicotinic acid 3-monooxygenase: A novel decarboxylative hydroxylase in bacterial nicotinate catabolism. *25th Enzyme Mechanisms Conference*, St. Pete Beach, FL, January **2017**.

<u>Katherine A. Hicks,</u> Meigan E. Yuen, Wei Feng Zhen, Anna M. Szostek, Tyler J. Gerwig*, Ryan W. Story*, Matthew R. Bauerle*, Alvi Sakib* and Mark J. Snider; Characterization of the catalytic mechanism of NicC, a 6-hydroxynicotinic acid 3-monooxygenase; *Gordon Research Conference on Enzymes, Coenzymes and Metabolic Pathways*, July **2016**.

<u>Mark J. Snider</u>; Structures and mechanisms of nicotinate catabolizing enzymes: A model system for investigating bacterial *N*-heterocyclic aromatic compound degradation and for undergraduate education. Central Eastern Regional Meeting of the American Chemical Society; Covington, KY, May **2016**.

<u>Nicholas P. Lesner</u>*, Michael T. Peterson, and Mark J. Snider (**2016**) Characterization and Identification of Metabolites in the Oxidation of Nicotinic Acid by *Bacillus Niacini*. *American Society of Biochemistry & Molecular Biology National Meeting*; **FASEB J 30:834.9**

<u>Morgan A Dasovich</u>*, Mark J Snider, and Michael T Peterson (**2016**) Characterization of a Novel Flavin-dependent Monooxygenase in *Bacillus niacini*. *American Society of Biochemistry & Molecular Biology National Meeting*; **FASEB J 30:834.8**

<u>Jessica A Meek*</u>, <u>Laura A Sherer</u>,* and Mark J Snider (**2016**) Deciphering the Metal Dependency of Novel Nicotinate Hydroxylase from *Bacillus niacini*. *American Society of Biochemistry & Molecular Biology National Meeting*; **FASEB J 30:1083.13**

Meigan E. Yuen, Weifeng Zhen, Megan C. Kopp, Tyler Gerwig*, Mark J. Snider, and <u>Katherine A. Hicks</u> (**2015**) Structural and Biochemical Characterization of *Pseudomonas putida* KT2440 NicC, a 6-Hydroxynicotinic Acid 3-Monooxygenase. Annual Meeting of the American Crystallographic Association.

<u>Tyler J Gerwig</u>*, Meigan Yuen, Weifeng Zhen, Mark A Anderson, Katherine Hicks and Mark J Snider (2015) Structure and mechanism of 6-hydroxynicotinate 3-monooxygenase (NicC). *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* 29:572.23.

<u>Abigail K Daniel</u>* and Mark J Snider (**2015**) Elucidating the nicotinic acid degradation pathway in *Bacillus niacini; American Society of Biochemistry & Molecular Biology National Meeting; FASEB J 29:573.19.*

<u>Lauren Buyan*</u> and Mark J Snider (**2015**) Virulence modulation of *Bordetella pertussis* via inhibition of the nicotinate degradation pathway. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 29:LB201.**

<u>Gentry J Kerwood</u>*, Manish Aryal*, Brittany Van Houten*, Dean M Fraga and Mark Snider (**2015**) Exploring the structural basis for negative cooperativity in the phosphagen kinase superfamily. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 29:724.12.**

<u>Karan Malani</u>* and Mark J. Snider (**2015**) Inhibition and kinetic isotope effect studies to further characterize the mechanism of maleamate amidohydrolase. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 29:572.13.**

- M. J. Snider, Z. Harvey*, H. Kondow*, P. Chiaranunt*, and M. Anderson; Mechanistic studies of nicotinate degrading enzymes and the identification of a novel nic cluster in *Bacillus niacini; Enzyme Structure and Function 37th Steenbock Symposium*, University of Wisconsin Madison, WI; May **2014.**
- <u>H. Kondow</u>*, and M. J. Snider (**2014**) Investigation of *B. bronchiseptica* maleamate amidohydrolase (NicF) substrate specificity. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **28:584.3.**
- M. Aryal*, and M. J. Snider (**2014**) Testing the versatility of the alternating sites of reactivity mechanism in the phosphagen kinases. *American Society of Biochemistry & Molecular Biology National Meeting;* FASEB J **28**:768.11. [Honorable Mention for Best Poster in DNA & Chromosomes Section of ASBMB National Undergraduate Poster Competition]
- <u>P. Chiaranunt</u>*, <u>J. Sprano</u>*, <u>A. Daniels</u>* and M. J. Snider (**2014**) Investigation of the nicotinate degradation pathway in *Bordetella pertussis* and its connections to whooping cough. *American Society of Biochemistry & Molecular Biology National Meeting;* **FASEB J 28:LB266**.
- Z. Harvey* and M. J. Snider (2014) Functional genomic elucidation of nicotinic acid catabolism in *Bacillus niacini*: Towards a metabolic engineering approach to environmental remediation. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* 28:980.2 [Honorable Mention for Best Poster in Bioenergetics Section of ASBMB National Undergraduate Poster Competition]
- <u>J. B. Claybourne</u>*, E. Shi*, S. Strand, M. Schultz and M. J. Snider (**2014**) Biodegradation of venlafaxine. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **28**:B289.
- M. M. Schultz, <u>M. J. Snider</u> and S. S. Strand (**2013**) Antidepressants contaminate your water! Investigations to determine whether bacteria can remediate. *Global Liberal Arts Alliance International Conference on Water*, The College of Wooster.
- <u>C. Young, M. J. Snider</u> and D. Fraga (**2013**) A curriculum based on research as pedagogy: A research project-based approach to teaching techniques in BCMB prepares students for senior independent study projects; *ASBMB Special Symposium on Student-Centered Education in the Molecular and Life Sciences*; Seattle University, Seattle, WA.
- M. J. Snider, P. Chiaranunt*, E. Sullivan*, K. Morrison, and D. Hilmey (2013) Investigating the catalytic role of Cys150-Asp29-Lys117 in the mechanism of maleamate amidohydrolase. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* 27:lb62.
- M. R. Bauerle*, W. Ammons*, K. Shvets* and M. J Snider (**2012**) Analysis of substrate specificity of 6-hydroxynicotinate-3-monoxygenase (NicC) from *Bordetella bronchiseptica*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **26**:963.13
- <u>S. Justice*</u> and M. J. Snider (**2012**) Investigation of the potential role of 6-hydroxynicotinate monooxygenase in the modulation of virulence in *Bordetella pertussis*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **26**:731.8
- <u>J. E. Noel</u>*, N. E. Spittle* and M. J. Snider (**2012**) On the role of Cys150 in the mechanism of maleamate amidohydrolase (NicF). *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **26**:756.20
- Brittany Begres*, A. Palmer*, J. V. Houten*, M. J. Snider, and D. Fraga (2012) Characterization of a

- hypotauracyamine kinase from the protozoan, *Phytophthora sojae*, and its implications on the evolution of substrate specificity in the phosphagen kinase family. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **26**:963.12
- M. J. Snider, M. Bauerle*, W. Ammons*, K. Shvets*, and E. Sullivan*; Aerobic catabolism of nicotinic acid (vitamin B₃) in *Bordetella bronchiseptica*: mechanistic studies of NicC and NicF; *Midwest Enzyme Chemistry Conference*, The University of Chicago; Oct. **2011**.
- <u>M. T. Henke*</u> and M. J. Snider; A mechanistic proposal for extradiol-like hydroquinone dioxygenases; *Midwest Enzyme Chemistry Conference*, The University of Chicago; Oct. **2011**.
- <u>J. Van Houten</u>*, A. Palmer*, K. Herring*, M. J. Snider, and D. Fraga (**2011**); Characterization of the *Phytophthora sojae* hypotaurocyamine kinase suggests early evolution of quaternary structure in the phosphagen kinase family. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:928.14
- M. J. Snider, B. A. Palanski*, Z. Rotter*, and X. Li (**2011**); Identification of the ligands of TM0486 from *Thermotoga maritima* by ESI-TOF mass spectrometry suggests role in a novel thiamin salvage pathway. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:923.4
- M. T. Henke* and M. J. Snider (**2011**); Mechanistic studies of 2,5-dihydroxypyridine 5,6-dioxygenase (NicX) from *Bordetella bronchiseptica*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:714.2
- <u>E. D. Sullivan*</u> and M. J. Snider (**2011**); Mechanistic studies of maleamate amidohydrolase (NicF) from *Bordetella bronchiseptica. American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:714.7
- R. Klein*, R. S. Rowlett, and M. J. Snider (**2011**); Structural studies of maleamate amidohydrolase (NicF) from *Bordetella bronchiseptica*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:714.4
- M. Bauerle*, R. Story*, and M. J. Snider (**2011**); Mechanistic studies of 6-hydroxynicotinate 3-monooxygenase (NicC) from *Bordetella bronchiseptica*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J* **25**:715.2
- M. J. Snider, B. A. Palanski*, A. Young* and Z. Rotter*; Guilt by association: functional annotation of TM0486 from *Thermatoga maritima* by identification of its bound ligands. *22nd Enzyme Mechanisms Conference*, St. Petersburg, FL, January **2011**.
- M. J. Snider, E. Sullivan*, M. Henke*, R. Klein*, M. Bauerle*, V. Kincaid*, R. Story* and R. Rowlett; Aerobic catabolism of nicontinic acid in Bordetella bronchiseptica: mechanistic studies of NicC, NicF and NicX. *22nd Enzyme Mechanisms Conference*, St. Petersburg, FL, January **2011**.
- <u>K. A. Stencel</u>*, M. Krasnor*, C. K. Koechli*, S. S. Strand, M. J. Snider and M. M. Schultz; Transformation of sertraline by aerobic sludge; *Proceedings of the 31*st *Society of Environmental Toxicology and Chemistry North American Meeting*, Portland, OR, November **2010**.
- E. A. Sakach*, M. J. Snider, H. L. Schoenfuss, M. M. Schultz; Accumulation of antidepressant pharmaceuticals in fish; *Proceedings of the 31st Society of Environmental Toxicology and Chemistry North American Meeting*, Portland, OR, November **2010**.

- <u>A.K. Palmer</u>*, D. Fraga, M. Snider and P. Edmiston (**2010**) Characterization of a dimeric arginine kinase in the protozoan, *Phytophthora sojae*, suggests an early origin of phosphagen kinase dimers. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **24**:469.7
- <u>V.A. Kincaid</u>* and M.J. Snider (**2010**) Nicotinic acid degradation in *Bordetella bronchiseptica*: mechanistic studies of the novel enzyme NicF. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J. 24:835.7 [Honorable Mention for Best Poster in Protein Section of ASBMB National Undergraduate Poster Competition]*
- <u>B.A. Palanski</u>* and M.J. Snider (**2010**) Guilt by association: functional annotation of TM0486 from *Thermotoga maritima* by identification of its bound ligands. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **24**:518.3 [Honorable Mention for Best Poster in Systems Biology Section of ASBMB National Undergraduate Poster Competition]
- M. J. Snider (2008) Negative cooperativity in transition state binding enhances rate of product release in phosphagen kinases; 40th Central Regional Meeting of the American Chemical Society; Columbus, OH.
- M. P. McGinley*, D. Fraga and M. Snider (2008) Evolution of cooperativity in the phosphagen kinase family. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* 22: 1004.3
- <u>V. M. Andrus</u>*, D. Thomas*, T. Meulia, M. Snider, and D. Fraga (**2008**) Characterization of the arginine kinase family of *Caenorhabditis elegans*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **22**: 1004.4
- <u>C. Clarke</u>*, and M. J. Snider (**2007**) Rate of spontaneous hydrolysis of creatine to estimate the catalytic proficiency of creatine amidinohydrolase. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **21**: 650.5
- <u>S. Agidi</u>*, and M. J. Snider (**2007**) Catalytic role of His232 in the mechanism of *Pseudomonas putida* creatine amidinohydrolase. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **21**: 805.2
- <u>D. H. Thomas*</u>, and M. J. Snider (**2007**) Investigating the physiological roles and intracellular localization of an arginine kinase in *Caenorhabditis elegans*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **21**: 510.11
- <u>J. Graham</u>*, and <u>L. Stetzik</u>*, M. J. Snider; Investigating cooperative ligand binding by rabbit muscle creatine kinase using isothermal titration calorimetry. *Midwest Enzyme Chemistry Conference*, Northwestern University, II. September **2006**.
- M. J. Snider, I. Lee, I. Eccles-James*, J. Anquandah*, J. Graham*, and L. Stetzik*; Dimerization enhances rate of product release in phosphagen kinase family. *Gordon Research Conference on Enzymes, Coenzymes and Metabolic Pathways*, University of New England, ME. July **2006**.
- <u>J. Anquandah</u>*, I. G. Eccles-James*, I. N. W. Lee, and M. J. Snider (**2006**) A mechanistic role for protein oligomerization in the phosphagen kinase family. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **20**: A44-a
- <u>E. E. Gustely</u>* and M. J. Snider (**2006**) Analysis of the role of His232 in the mechanism of Ps. putida creatinase. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **20**: A43-d

- <u>H.-D. Nguyen</u>*, and M. J. Snider (**2006**) Is the ATP analogue adenosine 5'-sulfatopyrophosphate an alternative substrate or inhibitor of creatine kinase? *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **20**: A43-b
- <u>B. D. Pipitone</u>*, and M. J. Snider (**2006**) Determining the subcellular localization of a novel arginine kinase in *Caenorhabditis elegans*. *American Society of Biochemistry & Molecular Biology National Meeting; FASEB J.* **20**: A900-b
- L. A. Horst*, Z.-M. Tun*, J. B. Tout*, M. J. Jourden*, J. A. Bayuk*, J. M. Shear*, <u>M. J. Snider</u>; Thermodynamic analysis of substrate binding and activation by rabbit muscle creatine kinase. *19*th *Enzyme Mechanisms Conference*, Pacific Grove, CA, January **2005**.
- M. R. Buckley*, V. Miraldi*, M. J. Snider; Rate of spontaneous phosphoryl transfer between ATP and creatine. 227th ACS National Meeting; Anaheim, CA, March 2004.
- <u>C. Chan</u>*, M. J. Snider, W. S. Kirk, W. W. Cleland; Primary ¹⁸O kinetic isotope effects on phosphoryl transfer by creatine kinase. *227th ACS National Meeting*; Anaheim, CA, March **2004**.
- <u>J. J. Ellinger</u>*, M. J. Snider, W. S. Kirk, W. W. Cleland; Primary ¹⁸O kinetic isotope effects for arginine kinase-catalyzed phosphoryl transfer from ATP. *227th ACS National Meeting*; Anaheim, CA, March **2004**.
- <u>D. E. Freeman</u>*, M. J. Snider; Is creatine kinase catalytically promiscuous? *227th ACS National Meeting*; Anaheim, CA, March **2004**.
- J. E. Hall*, M. J. Snider; Biochemical characterization of a Caenorhabditis elegans arginine kinase. 227th ACS National Meeting; Anaheim, CA, March 2004.
- R. Wolfenden, M. J. Snider; A transition state analogue complex in the vapor phase; 18th Enzyme Mechanisms Conference; Galveston Island, TX, January **2003**.
- M. Borders, M. J. Snider, Geiss, P.R.*, Jourden, M.J.*, Kersteen, E.A.*, Moore, N.R.*, McBride, L.C.*, Rohan, M.C.*, Almeida, R.M.*, Gbeddy, E.R.K.*, Mulligan, G.B.*, MacGregor, K.M.*, Lake, B.A.*, Bohl, J.L.*, Terrano, D.T.*, Edmiston, P.L.; Creatine kinase is more than positive about it: roles of each component of a six-arginine cluster at the active site in binding and catalysis; 18th Enzyme Mechanisms Conference; Galveston Island, TX, January 2003.
- <u>J. M. Rackley</u>*, C. L. Borders, Jr., M. J. Snider; Temperature dependence of the catalytic activity of rabbit muscle creatine kinase; *16th National Conference on Undergraduate Research*, University of Wisconsin Whitewater, WI, April **2002**.
- <u>D. R. Korstjens</u>*, M. J. Snider, C. L. Borders, Jr.; Renaturation of insoluble mutants of creatine kinase; 16th National Conference on Undergraduate Research, University of Wisconsin Whitewater, WI, April **2002**.
- <u>R. Wolfenden</u>, Mark J. Snider; Site-bound water and the limitations of a potential transition state analogue inhibitor; *Gordon Research Conference on Enzymes, Coenzymes and Metabolic Pathways*, Meriden, NH, July **2001**.
- <u>M. J. Snider</u>, R. Wolfenden; Thermodynamic role of site bound water in transition state and transition state analogue binding by cytidine deaminase; *17*th *Enzyme Mechanisms Conference*; Marco Island, FL, January **2001**.

M. J. Snider, S. Gaunitz, C. Ridgway, R. Wolfenden; Temperature dependence of the transition state affinity of cytidine deaminase; *219*th ACS National Meeting; San Francisco, CA, March **2000**.

INVITED RESEARCH TALKS

2017	Dept. of Chemistry, SUNY-Buffalo
2016	Central Regional American Chemical Society Meeting, Covington, KY.
2013	Natural Science Division, Rio Grande University, OH
2012	Dept. of Chemistry, University of Toledo, OH
2011	Dept. of Chemistry & Biochemistry, Denison University, OH
	Dept. of Physiology and Biophysics, Case University, Cleveland, OH
2009	Dept. of Biochemistry & Biophysics, U. North Carolina - Chapel Hill
2008	Dept. of Chemistry, Capital University
	40th Central Regional Meeting of the American Chemical Society;
	Symposium in honor of Prof. Ming-Daw Tsai; Columbus, OH
	Dept. of Chemistry, Ithaca College, Ithaca NY
	Dept. of Biol. & Chem. Sciences, Wells College, Aurora, NY
2007	Dept. of Chemistry, The Ohio State Univ., Columbus, OH
2006	Dept. of Biochemistry & Molecular Biol., Pennsylvania State Univ.
	Dept. of Chemistry, The Univ. of Akron
	Begley Laboratory, Cornell Univ.
2005	Dept. of Chemistry, Michigan State Univ.
	Dept. of Biochemistry & Biophysics, Univ. North Carolina – C.H.
2004	The Wooster Section of the American Chemical Society; at Ashland Univ.
2003	Dept. of Medicinal Chem & Mol. Pharm., Purdue Univ.
	Dept. of Chemistry, Capital Univ.
2002	Dept. of Chemistry, Beloit College
	Dept. of Chemistry, John Carroll Univ.
	Dept. of Chemistry, Middlebury College
	Wooster Area Molecular Biology Association, OARDC, OSU
2001	Structural Biology & Biotechnology Symposium, Chapel Hill, NC

PROFESSIONAL MEMBERSHIPS

American Society for Biochemistry & Molecular Biology 2005– American Chemical Society: Biological Division & Chemical Education 1993– Council on Undergraduate Research 1999–2007

COURSES TAUGHT AT THE COLLEGE OF WOOSTER (*course taught regularly)

Controversies in Science and Public Policy (Chemistry 103; chemistry course for non-science majors) Chemistry Tutorial: The Life and Death of a Molecule (CHEM 400)

First-Year Seminar in Critical Inquiry (Interdepartmental writing intensive course for incoming students)

- *General Chemistry I (CHEM 111)
- *General Chemistry II and Laboratory (CHEM 112)

Organic Chemistry Laboratory (CHEM 211L and 212L)

Organic Chemistry II (CHEM 212)

- *Techniques in Biochemistry & Molecular Biology (BCMB 303)
- *Principles of Biochemistry (BCMB 331)
- *Biochemistry of Metabolism (BCMB 332)
- *Biophysical Chemistry (CHEM 334)

Advanced Biochemistry Tutorials (BCMB 400)

- *Introduction to Independent Study (BCMB 401)
- *Senior Independent Study (CHEM & BCMB 451/452)

ELECTED AND APPOINTED COMMITTEE SERVICE AT THE COLLEGE OF WOOSTER

ELECTED AND APPOINTED COMMINITIES SERVICE AT THE COLLEGE OF WOO	SIER
Biochemistry & Molecular Biology Curriculum Committee,	2001-present
Chairperson	2008-2011; 2020-2023; 2025-
College of Wooster Scholarships at State Science Day,	2001-2007; 2009-2010; 2014
Judging Chairperson	2001-2007
College Scholars Examination Committee	2001-2004
Conference with Trustees Committee (elected)	2020-2022
Department of Chemistry Chairperson	2013-2016
EPC Sub-Committee for Assessing Quantitative Reasoning Courses	2006
Faculty Development Committee	2008-2009
Faculty Research & Study Leaves Committee	2008-2009
Financial Advisory Committee (elected)	2004-2006
Five-Year Strategic Planning, Priorities Advisory Committee (elected)	2004
Henry J. Copeland Funds for Independent Study Committee	2002-2004; 2006-2007
Howard Hughes Medical Institute Steering Committee	2011-2012
Interfaith Campus Ministries Advisory Board	2003-2007
Life Sciences Planning Committee and Ruth W. Williams Hall of Life Sciences	ence Facility 2011-2018
Co-Building Shepherd (with Dr. Dean Fraga)	2013-2018
Pre-Health Advisory Committee	2003-2007
Teaching Staff and Tenure Committee (elected)	2009-2012; 2018; 2023-2026
Co-Chair (with Provost)	2011-2012; 2023-2025
Upperclassman Programs Committee	2003-2004
Wellness Program Committee	2006-2007
SCHOLARLY COMMUNITY SERVICE	
Middle School Science Fair Judge at St. Mary Elementary School, Wooster	2018-2020; 2023-2025
Scientific Judge at the ASBMB National Undergraduate Research Poster Com	
NSF Grant Proposal Review Panel	2017; 2021; 2023
Curriculum Reviewer for Department of Chemistry, Elon University	April 2016
Preparing Future Faculty (Ohio State University) Mentor	2009 – 2010
Instructor, B-WISER Summer Camp for 8 th Grade Girls	2006; 2019; 2022
Chemistry Faculty Mentor, Council of Undergraduate Research	2006 – 2008
Technical Expert, Wayne County Local Emergency Planning Committee	2011– 2013
PEDAGOGICAL DEVELOPMENT	
Inclusive Research Mentoring Faculty Learning Community, College of Woo	
KinTek New Enzymology Kinetics Workshop; Brno, Czech Republic	May 2019
ASBMB Regional Workshop at The College of Wooster (co-hosted with Dea	
Implementing Vision and Change Using Concept-Driven Teaching Strategie	
ASBMB Regional Workshop at Kentucky Wesleyan University, Owensboro,	•
Implementing Vision and Change Using Concept-Driven Teaching Strategie	
ASBMB Special Symposium on Student-Centered Education in the Molecula	_
Sciences; Seattle, WA. Poster presentation entitled, with Crystal Young: A	
on research as pedagogy: A research project-based approach to teaching	techniques
in BCMB prepares students for senior independent study projects	
Wooster Faculty Workshop: First-Year Seminars in Critical Inquiry	May 2011
POGIL Workshop; Washington College; Chestertown, MD	June 2008

American Academy of Colleges & Universities: The importance of institutional, disciplinary, and interdisciplinary definitions of scholarship; The Student as Scholar: Undergraduate Research and Creative Practice; Long Beach, CA. April 2007 Hewlett-Mellon Presidential Discretionary Fund for Institutional Renewal: A Comprehensive Examination of Introductory Science and Mathematics Courses 2006-2007 Five Colleges of Ohio Assessment Conference, Wooster, Ohio February 2006 Project Kaleidoscope National Colloquium: Translating How People Learn into a Roadmap for Institutional Transformation; Kansas City, Missouri October 2005 Wooster Faculty Workshop: First-Year Seminars in Critical Inquiry May 2004 Wooster Faculty Workshop on Teaching and Learning: "How Well Are Students Learning What We're Teaching?" with Thomas A. Angelo February 2004 Project Kaleidoscope Assembly: Motivating Students to Pursue Careers in STEM Fields; Oberlin College, Oberlin, Ohio September 2003 Wooster Faculty Workshop on Writing in the Sciences with J. Pechenik April 2003 Wooster Faculty Conference on Teaching and Learning May 2002

SENIOR INDEPENDENT STUDY THESIS RESEARCH ADVISEES (96 graduates)

#	I.S. Advisee	Grad Year	Major	Thesis Title	Post-Wooster
1	Davin R. Korstjens	2002	Biochemistry	The attempted refolding of the insoluble rabbit muscle	M.S. (2005) in BioMedical
				creatine kinase R214K mutant into an active enzyme.	Sciences, Univ. of Colorado
2	Jane M. Rackley	2002	Chemistry	The temperature dependence of the catalytic activity of	M.Ed. Carlow Univ. (2012);
				rabbit muscle creatine kinase.	H.S. Chemistry Teacher
3	Jennifer A. Bayuk	2003	Biochemistry	Temperature effects on substrate binding and activation	D.O. (2008) Kirksville College
				by creatine kinase.	of Osteopathic Medicine
4	Virginia A. Miraldi	2003	Biochemistry	Development of a quantitative protocol using ³¹ P NMR	M.D. (2007) CWRU;
				spectroscopy to study the reaction of the spontaneous	Assistant Professor,
				phosphoryl transfer from MgATP to creatine.	Department of
					Ophthalmology, U.
5	Tabetha D. Shelly	2003	Diachamietm	The expression and purification of a <i>C. elegans</i> arginine	Cincinnati Research Scientist, R&D,
5	Tabetha D. Shelly	2003	Biochemistry	kinase.	Leiner Health Products, NC;
				Killase.	Registered Nurse at
					Piedmont Medical Center
6	Marshall R. Buckley	2004	ВСМВ	Progress toward measuring the rate of spontaneous	MBA, Boston College (2008)]
		200.	202	phosphoryl transfer from MgATP to creatine.	
7	Chikio Chan	2004	ВСМВ	Progress toward measuring primary ¹⁸ O kinetic isotope	Ph.D., Biochemistry, Yale
				effects on phosphate transfer by creatine kinase.	University (2010); Asset
					Management, Colorado
8	James J. Ellinger	2004	ВСМВ	Determining the rate-limiting step of the reaction	Ph.D., Biochemistry, Univ. of
				catalyzed by Stichopus japonicus arginine kinase by	Wisconsin (2012); Assistant
				viscosity variation.	Professor, The University of
					Tokyo, Japan
9	Dena E. Freeman	2004	ВСМВ	Is creatine kinase catalytically promiscuous?: synthesis,	MPH, Univ. of Washington
				purification and kinetic analysis of adenosine 5'-	(2008)
				sulfatopyrophosphate as a potential substrate for	
				creatine kinase.	
10	Joseph E. Hall	2004	ВСМВ	Biochemical characterization of a novel Caenorhabditis	M.D., Univ. of Cincinnati
				elegans arginine kinase through linked-enzyme kinetic	(2008); ENT Fellow,
				assay analysis.	Vanderbilt Univ.; Practicing
11	linema C. Faalaa	2005	DCMD	Investigation a satelytic value of discovering for a set of	ENT in Columbus, OH
11	Ijeoma G. Eccles-	2005	ВСМВ	Investigating a catalytic role of dimerization for creatine	M.Sc. in Immunology,
	James			kinase.	Imperial College, London, UK (2007); Research
					Associate, UC-SF
12	Jennifer L. Shrock	2005	ВСМВ	Exploring the amino acid determinants of cooperative	BSN, Regis University (2010)
12	Jennier L. Jillock	2003	DCIVID	transition state binding by creatine kinase.	Doir, Negis Offiversity (2010)
13	Jason B. Tout	2005	Chemistry	Thermodynamic changes accompanying the binding of	M.Ed., Boston College
13	133011 2. 1041	1 2003	Sileniisti y	MgADP in a composite, quaternary transition state	(2008); High school science
				analogue complex by creatine kinase.	teacher
14	Lauren M. Wagner	2005	ВСМВ	Exploring the cellular localization of a novel arginine	Ph.D., Molecular Biology,
			-	kinase in <i>Caenorhabditis elegans</i> by fluorescence	Univ. of Pittsburgh (2012)
				microscopy.	
15	Juliana Anquandah	2006	ВСМВ	Investigating a mechanistic role for protein	M.D., St. George's
	,			oligomerization in the phosphagen kinase family.	University, Grenada, West
		•		1.5	• •

					Indies (2013); Obstetrics-
					Gynecology Residency
16	Erin E. Gustely	2006	ВСМВ	Investigating the mechanism of creatinase	M.S., Biochemistry, Yale University (2007); High school science teacher
17	Hai Dang Nguyen	2006	ВСМВ	ATP analogue, adenosine 5'-sulfatopyrophosphate: an alternative substrate or inhibitor of creatine kinase?	Ph.D., Biochemistry, Univ. of Minnesota (2012); Post- doctoral Fellow at Harvard Medical School; Assistant Professor, Department of Pharmacology, Univ. of Minnesota (2019-)
18	Baldassare Daniel Pipitone	2006	ВСМВ	Examining the physiological role of an arginine kinase in Caenorhabditis elegans.	D.O., Ohio University (2010); Neuro-radiology Fellow, Univ. Michigan (2017); practicing radiologist
19	Perfect Senyo Agidi	2007	ВСМВ	Kinetic mechanism of <i>Pseudomonas putida</i> creatine Amidinohydrolase by isothermal titration calorimetry.	D.O. (2015) Nova Southeastern University, Florida; Internal Medicine Specialist, Akron
20	Chafen Clarke	2007	ВСМВ	Catalytic proficiency of creatine amidinohydrolase	DDS (2015) The University of the West Indies, Jamaica
21	David H. Thomas	2007	ВСМВ	Determining the sub-cellular localization of an arginine kinase in <i>Caenorhabditis elegans</i> .	M.D., Univ. of Cincinnati (2012); Internal Medicine, Case Univ. Hospital
22	Christopher Chapman	2009	Chemistry	Kinetic analysis of spontaneous creatine hydrolysis by ¹ H NMR and creatine amindinohydrolase catalyzed creatine hydrolysis by isothermal titration calorimetry.	Chemist at PPG Industries, Inc.
23	Stephanie Edmisson	2009	BCMB	Investigating the role of the IQ67 domain in the interaction between SUN and calmodulin: an endeavor into the treacherous world of protein expression by <i>Eschericia coli</i> (under the direction of Dr. Esther van der Knaap, OARDC)	High School Biology Teacher
24	Theodore Moore III	2009	ВСМВ	Characterization of a putative monooxygenase involved in NAD catabolism	Ph.D., (2015) Microbiology, Univ. Wisconsin-Madison; Post-doctoral Fellow, MIT; Program Lead, Gingko Bioworks, Cambridge MA.
25	Allyson Palmer	2010	ВСМВ	Expression and characterization of <i>Phytophthora sojae</i> phosphagen kinases	M.D. Ph.D. (2018), The Mayo Clinic, MN
26	Ryan W. Story	2010	ВСМВ	NAD catabolism: identification of a 6-hydroxynicotinate 3-monooxygenase in <i>Bordetella bronchispectica</i>	M.D. (2014) The Ohio State University; Residency, Vanderbilt Univ.; Practicing pulmonologist
27	Virginia Kincaid	2010	Chemistry	Nicotinic acid degradation in <i>Bordetella bronchiseptica</i> : kinetic studies of the hydrolytic deamidase NicF.	Ph.D. Biochemistry (2017) Univ. of Wisconsin – Madison; Senior Research Scientist, Promega Corp.
28	David Flannelly	2010	Chemistry	Wastewater, the new lipstick in feminization: the analytical determination of a range of endocrine disruptors in the Chicago watershed.	Teach for America Program; M.S. Environmental Toxicology/Chemistry, Cornell University; High School Science Teacher
29	Roger D. Klein	2011	A.C.S Certified Biochemistry, BCMB, & Physics	A structure-function analysis of maleamic acid amidinohydrolase (NicF) from <i>Bordetella bronchiseptica</i>	M.D. Ph.D. (2020), Washington University, St. Louis; Residency in Urology, Univ. Pittsburgh
30	Matthew T. Henke	2011	всмв	Developing a mechanistic proposal for 2,5-dihydroxypyridine-5,6-dioxygenase (NicX) from Bordetella bronchiseptica.	Ph.D. (2016) Integrative Biological Sciences, Northwestern University; Post-Doctoral Fellow, Harvard University; Assistant Professor of Pharmacology, University of Illinois at Chicago (2022)
31	Brad A. Palanski	2011	A.C.S. – certified Biochemistry; BCMB	An investigation of the ability of <i>Pseudomonas</i> fluorescens to degrade the antidepressant sertraline (Zoloft®)	Ph.D. (2018), Chemistry, Stanford University; Post- Doctoral Fellow, Harvard Medical School; Director of Biochemistry,

					Vaccine Company Inc., San Francisco, CA (2022)
32	Eric D. Sullivan	2011	всмв	Mechanistic studies of maleamate amidohydrolase (NicF) from <i>Bordetella bronchiseptica RB50</i> .	Ph.D. (2016), Chemical Biology, University of Michigan; Post-doctoral Fellow, NIEHS, Raleigh, NC
33	Matthew R. Bauerle	2012	A.C.S certified Chemistry	Structure-function analysis of 6-hydroxynicotinate 3-monooxygenase.	Ph.D. (2018), Chemistry, Pennsylvania State University; Senior Research Chemist, Exemplify Biopharma, Inc.
34	Jonathon Fox	2012	ВСМВ	Isolation of bacterial strains in wastewater sludge capable of degradation of sertraline, venlafaxine and triclocarban.	MS (2017), Biomedical Engineering, Cleveland State University; Engineer at MED Institute, West Lafayette, IN
35	Samantha Justice	2012	ВСМВ	Investigation of the potential role of 6- hydroxynicotinate-3-monooxygenase in the modulation of virulence in <i>Bordetella pertussis</i>	Ph.D. student, Biological and Biomedical Sciences, Univ. Southern California
36	Jeffrey E. Noel	2012	ВСМВ	Investigation in the catalytic mechanism of maleamate amidohydrolase (NicF) from <i>Bordetella bronchiseptica</i> RB50	unknown
37	Nicholas E. Spittle	2012	ВСМВ	Probing the role of cysteine-150 in maleamate amidohydrolase (NicF) catalysis from <i>Bordetella bronchiseptica RB50</i> by site-directed mutagenesis.	D.O. (2016) Ohio University College of Osteopathic Medicine; Orthopedic surgeon
38	Zachary Harvey	2014	A.C.S-certified Biochemistry	Defining the Genome and <i>nic</i> Cluster of <i>Bacillus niacini</i> .	Ph.D. (2019) Chemical & Systems Biology, Stanford University; Post Doctoral Fellow at the Gregor Mendel Institute, Vienna, Austria
39	Manish Aryal	2014	ВСМВ	Evolution of negative cooperativity in phosphagen kinases	Ph.D. student, Molecular Biophysics and Structural Biology Program, Carnegie Mellon / Univ. Pittsburgh
40	Pailin Chiaranunt	2014	BCMB & Philosophy	A scientific paradigm shift from reductionism to holism: the potential relationship between nicotinate degradation and virulence modulation in <i>Bordetella</i> pertussis	Ph.D. student, Immunology, University of Toronto, Canada
41	Helena Kondow	2014	ВСМВ	Elucidation of <i>Bordetella bronchiseptica</i> maleamate amidohydrolase (NicF) substrate specificity	Ph.D. (2020) Biochemistry & Biophysics, Texas A&M University; ORISE Fellow; Business Development Manager, Gainsville, VA.
42	Jacob Sprano	2014	BCMB & History	Pertussis, pellagra, and poverty: a historical and scientific analysis of whooping cough's associations with poverty and pellagra, and the nicotinate degradation pathway in <i>Bordetella pertussis</i>	M.D. (2019), Kansas City University of Medicine; Resident in Family and Community Medicine, Univ. of Nevada, Reno, NV
43	James Claybourne	2014	Neuroscience/ BCMB	An analysis of the bioremediation of venlafaxine using aerobic bacteria	applying to law school
44	Lauren Buyan	2015	Neuroscience/ BCMB	Virulence modulation of Bordetella pertussis via inhibition of the nicotinate degradation pathway	D.O. (2019), College of Osteopathic Medicine of the Pacific Northwest, Oregon; Residency in Emergency Medicine, Univ of Nebraska Med Center
45	Abigail Daniel	2015	ВСМВ	Cloning and expression of the novel proteins in the Bacillus niacin nicotinic acid degradation pathway and an initial attempt to characterize the NicAB complex	Apprenticeship in sustainable agriculture, Philadelphia, PA; High school biology teacher
46	Tyler Gerwig	2015	Chemistry	Assessing mechanistic proposals of 6-hydroxynicotinate- 3-monooxygenase (NicC) from <i>B. bronchiseptica</i> .	D.O. (2020) Ohio University College of Osteopathic Medicine; Internal Medicine Residency, US Naval Medical Center, Portsmouth, VA.
47	Karan Melani	2015	ВСМВ	Inhibition and kinetic isotope effect studies to further characterize the mechanism of maleamate amidohydrolase (NicF).	D.P.M. (2019), Temple University; Practicing in Naples, FL
48	Alvi Sakib	2016	ВСМВ	Investigating the proposed acid-base catalyzed mechanism for 6-hydroxynicotinate 3-monooxygenase (NicC)	M.B.A. (2018) University of Michigan; Business Consultant, Deloitte,

					Chicago, IL
49	Morgan Dasovich	2016	ВСМВ	The partial characterization of a monooxygenase in Bacillus niacini	Ph.D. (2023), Chemical Biology, Johns Hopkins University; Post-doctoral Fellow at Johns Hopkins Univ. and UT-SWMC
50	Melia Kovach	2016	ВСМВ	Generation of a ΔnicC Bordetella pertussis mutant: investigating the relationship between nicotinic acid degradation and virulence modulation.	Physician Assistant / MPH (2020); Northeastern University. Practicing PA in Boston, MA.
51	Nicholas Lesner	2016	A.C.S certified Chemistry	Attempted synthesis of 2,3,6-trihydroxypyridine and characterization of the putative ring-cleavage enzyme in <i>Bacillus niacini</i> nicotinate catabolism	Ph.D. (2021), Biochemistry and Biophysics, Univ. of Texas – Southwest Medical Center; Post-doctoral Fellow, Univ. Penn.
52	Dylan McCreary	2016	A.C.S certified Chemistry	Investigation of the functionality of the nicotinic acid degradation pathway of <i>Bordetella pertussis</i> as a potential treatment of whooping cough	M.D. (2020), Univ. of Minnesota Medical School; Residency in Internal Medicine, Univ of Arizona
53	Jessica Meek	2016	ВСМВ	Deciphering the metal dependency of novel nicotinic acid dehydrogenases from <i>Bacillus niacini</i>	M.D. (2020), Univ. College Cork School of Medicine, Ireland; Residency in Internal Medicine, Brown University.
54	Jack Harrington	2017	Chemistry	Elucidating the mechanism of oxidative decarboxylation of 6-hydroxynicotinate by 6-hydroxynicotinate 3-monooxygenase (NicC) through kinetic analysis	Taught English in China; Industrial research chemist
55	Kathleen Kalafatis	2017	Neuroscience/ BCMB	An attempt to characterize the multi-subunit nicotinic acid and 6-hydroxynicotinic acid dehydrogenase complexes in the nicotinic acid degradation pathway of <i>Bacillus niacini</i>	D.O. student, Lincoln Memorial University DeBusk College of Osteopathic Medicine
56	Laura Sherer	2017	ВСМВ	Biophysical characterization of the cofactors in the putative NicAB complex from <i>Bacillus niacini</i>	Ph.D. (2022) Univ. of Minnesota
57	Tsun Ki Jerrick To	2017	ВСМВ	Characterizing the genetic determinants of the nicotinic acid degradation pathway in <i>Bacillus niacini</i>	Ph.D. (2022), Univ. of Pennsylvania; Consultant with Boston Consulting Group
58	Nathan Brownstein	2018	ВСМВ	Attempt at expression of a functional recombinant nicotinate dehydrogenase from <i>Bacillus niacini</i>	Ph.D. (2025) Brandeis Univ.
59	Sydney Fine	2018	ВСМВ	Niacini, now you don't: the catabolism of nicotinic acid and nicotine and the role of a putative monooxygenase enzyme	Research Intern, Fred Hutchinson Cancer Center, Seattle, WA; MPH student, Global Health Epidemiology Program, The University of Michigan
60	Kent Nakamoto	2018	A.C.S certified Chemistry	Elucidating the mechanism of 6-hydroxynicotinate 3-monooxygenase, an enzyme involved in aerobic degradation of nicotinate	M.D. (2022) The Ohio State University
61	Derek Schwarz	2018	ВСМВ	Help! Help! I'm being repressed!: An in-depth analysis of the IcIR and its in vitro oligomerization	Ph.D. student, University of Akron
On leave	Ryan Campbell	2019	ВСМВ	A Mechanistic Investigation: Probing the Substrate Promiscuity of 6-hydroxynicotinic acid 3-monooxygenase (NicC) from Bordetella bronchiseptica RB50	Post-Baccalaureate Research, NIH; Ph.D. student, Dartmouth
On leave	Kaeli Zoretich	2019	ВСМВ	Progress Towards Understanding Nicotinic Acid Degradation and the nic Operon in B. niacini Using Differential Gene Expression Analysis	M.D. (2023), The Ohio State University
62	Anh Dinh	2020	ВСМВ	Differential expression analysis of the <i>nic</i> cluster and attempts at characterizing the NicAB complexes in novel nicotinic acid degradation pathway of <i>Bacillus niacini</i>	Ph.D. student in Biochemistry, Virginia Tech
63	Noah Helton	2020	ВСМВ	Progress in characterizing a novel root exudate: differential gene expression of <i>Bacillus niacini DMS 2993</i> in response to nicotinic acid through RNAseq	Ph.D. student in Genetics, University of Michigan
64	Brent Pedersen	2020	ВСМВ	Swapping the <i>N</i> -termini of mitochondrial and cytoplasmic taurocyamine kinase from <i>Arenicola brasiliensis</i> in order to study the importance of the interactions between the N and C termini in controlling substrate binding and specificity	applying to medical school
65	Scott Perkins	2020	Neuroscience/ BCMB	The binding and kinetic mechanisms of 6-hydroxynicotinate 3-monooxygenase (NicC), a decarboxylative hydroxylase involved in nicotinate degradation	M.D. student, Cleveland Clinic Lerner College of Medicine, CWRU

66	Phillip Plotkin	2020	Neuroscience/ BCMB	Determining the roles of arginine 184 and arginine 108 of 6-hydroxynicotinic acid 3-monooxygenase (NicC) from <i>Bordetella bronchiseptica RB50</i> in substrate binding.	
67	Anneliese Wagoner	2020	ВСМВ	Moorea than you bargained for: the attempted confirmation and characterization of a novel arginine kinase in Moorea producens	Ph.D. student in Molecular & Cellular Biology, Ohio University
68	Luke Borgelt	2021	ВСМВ	Reconstitution of hydroxylation activity of recombinant NDH and 6-HDH complexes from <i>Bacillus niacini</i>	Researcher at Boston Hospital; MD student, Univ of Toledo
69	Emmalee Cooke	2021	ВСМВ	Identification of substrate specific arginine kinases and Taurocyamine kinase in the study of phosphate kinase evolution within flatworms.	Researcher at CWRU, then applying to medical school
70	Daniel Fleming	2021	ВСМВ	Establishing a functional heterologous expression system for <i>Bacillus niacini</i> nicotinate dehydrogenase and 6-hydroxynicotinate dehydrogenase using <i>E. coli</i>	DDS (2025), The Ohio State University
71	Wenhao "Martin" Hu	2021	ВСМВ	Homology modeling and protein-ligand structural analysis of the <i>Bacillus niacini</i> NicAB protein complex	M.S. (2023) Florida State Univ.; PhD student at University College London
72	Mingyuang "Merlin" Li	2021	ВСМВ	Evidence of negative cooperativity in cytosolic Taurocyamine kinase from Arenicola brasiliensis and its implication in the evolution of the phosphagen kinase family	Ph.D. student, Johns Hopkins Univ.
73	Connor Mangan	2021	ВСМВ	Elucidating the biodegradative pathway of nicotinic acid by <i>Bacillus niacini</i> : defining the biochemical action of <i>Bn</i> DUF	M.D. student, Uniformed Services University of Health Sciences, Bethesda
74	Jack Redick	2021	ВСМВ	Characterization of substrate promiscuity for 6-hydroxynicotinate-3-monooxygenase: a transient kinetic investigation of the decarboxylative-hydroxylation of 5-Cl-6-hydroxynicotinic acid	M.D. (2025), The Ohio State University; Internal Medicine Residency, OSU
75	Robert Beal	2022	ВСМВ	Role of tight binding in negative cooperativity in the dimeric mitochondrial taurocyamine kinase from Arenicola brasiliensis	PhD student, University of Cincinnati
76	Joel Brown	2022	ВСМВ	Molecular recognition within the ES complex: a mechanistic investigation to discern how 6-hydroxynicotinate-3-monooxygenase (NicC) distinguishes between substrate analogues when enhancing its rate of NADH oxidation	Sutra Biopharma, San Francisco; Ph.D. student, University of Michigan
77	Emilee Haines	2022	ВСМВ	Advances in characterizing the gene-encoded subunits that compose the nicotinate dehydrogenase and 6-hydroxynicotinate dehydrogenase enzyme complexes within <i>Bacillus niacini</i> nicotinic acid degradation pathway	Ph.D. student, Kent State University
78	May Z. Hlaing	2022	ВСМВ	Investigating substrate specificity for 6-hydroxynicotinate-3-monooxygenase (NicC) with alternative substrates: consequences to catalysis of replacing or removing the ring nitrogen	Biomedical researcher, The James Cancer Center at OSU; M.D. student, UCLA
79	Sai Kwan Khal	2022	ВСМВ	Advances in the elucidation of the nicotinic acid degradation pathway in <i>Bacillus niacini</i> : proof of formation of the trihydroxypyridine intermediate and the characterization of an FMN-dependent flavoprotein of unknown function	PhD student, Duke University
80	Omar Kelly	Dec 2022	ВСМВ	The Influence of Key Residues on Catalytic Activity and Substrate Specificity of Taurocyamine Kinases from Arenicola brasiliensis, An enzyme in the Phosphagen kinases family	MS student, Georgia State University
81	Eric Adadevoh	2023	ВСМВ	Exploring the Binding Multiplicity, Mode of Inhibition and Communication between NADH Binding Sites on 6-Hydroxynicotinate 3-monooxygenase (NicC)	Research position Ruby Biotech, San Francisco; M.S. (2025) in Sustainable Business, Univ. Miami
82	Sam Belsky	2023	ВСМВ	Interrogation of a conserved overlap between cofactor and substrate binding domains to facilitate crosstalk-induced conformational changes in Class A Flavin Monooxygenases: A case study of 6-Hydroxynicotinate-3-Monooxygenase	PhD student, Univ. Illinois – Urbana Champaign
83	Kira Boyce	2023	BCMB	A CoxG homologue subunit is necessary for the function of <i>Bacillus niacin</i> nicotinate dehydrogenase	PhD student, University of Virginia
84	Takoda Zuehlke	2023	ВСМВ	Identification of a putative binding mechanism for the reducing substrate of 6-hydroxynicotinate 3-monooxygenase	PhD student, University of Pittsburgh; Oncology
85	Jack Donahue	2024	ВСМВ	Role of residues R184 and W273 in 6-HNA and NADH binding by 6-hydroxynicotinate 3-monoxygenase	M.D. student; Pennsylvania State University Medical School

86	Grace Gascoigne	2024	ВСМВ	Structural and kinetic analysis of <i>Perkinsus Marinus</i> creatine kinase: insight to quaternary structure evolution of phosphagen kinases	M.D. student; University of Cincinnati College of Medicine
87	Abigail Lyon	2024	ВСМВ	Elucidating the nicotinic acid degradation pathway in Bacillus niacini: investigating the catabolic function of a flavin monooxygenase (FMO), domain of unknown function (DUF), and hypothetical protein (HP)	Research Tech; plans to apply to graduate school
88	Willow Reddish	2024	ВСМВ	Investigating activity loss in an engineered monomeric creatine kinase through the lens of a sponge-derived CK — an exploration of quaternary structure	D.D.S. student, Case Western Reserve University
89	Kath Olson	2024	ВСМВ	Determining the role of FMO in the <i>Bacillus niacini</i> nicotinic acid degradation pathway	Ph.D. student, The Ohio State University
90	Jorge Virto	2024	ВСМВ	Insights into pyocyanin synthesis: unraveling the enzymatic interplay of PhzS and PhzM from Pseudomonas aeruginosa Boston 41501 in vitro	Research Tech at CWRU; Plans to apply to graduate school
91	Aglesia Aregawi	2025	ВСМВ	Investigating substrate promiscuity of 6- hydroxynicotinate 3-monooxygenase (NicC): NicC activity with 5-chloro-coumalate	
92	Julian Pac	2025	ВСМВ	Deciphering pyocyanin biosynthesis: Investigating PhzS/PhzM enzyme dynamics and substrate interactions	
93	Mayank Pandey	2025	ВСМВ	Investigating the importance of Arg 108 in the binding of NADH by 6-hydroxynicotinate 3-monooxygenase (NicC)	Research Tech; plans to apply to graduate school
94	Charitha Patlolla	2025	ВСМВ	Roles of Arg184 and His302 in substrate binding and coenzyme coordination by 6-hydroxynicotinate 3-monooxygenase (nicc)	M.S. student, University of Chicago; plans to apply to Law School
95	Grant Rivera	2025	ВСМВ	Investigating the auxiliary site of 6-hydroxynicotinate 3- monooxygenase: kinetic characterization of the N354L and N354D variants	Applying to Medical School
96	Sipara Semu	2025	ВСМВ	Investigation into the impact of R108M and R184K variants of 6-hydroxynicotinate-3-monooxygenase on 6-HNA binding and flavin mobility	M.S. student, Northwestern University; applying to Medical School
97	Charlie Cuttino	2026	ВСМВ		
98	Jake Enzman	2026	ВСМВ		
99	Seongmin Ro	2026	ВСМВ		
100	Abbigail Williams	2026	всмв		
101	Caley Sherril	2026	ВСМВ		